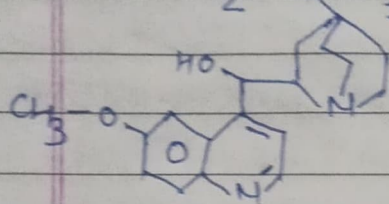




Isolation, Identification & Analysis of phytoconstituents ## Alkaloids #CH₂=CH # Quinine # its structure-ii) Chemo-taxonomic distribution-

- Quinine & quinidine isolated from Cinchona bark.
- Cinchona is the dried bark stem & root is used for the synthesis of quinine & quinidine-
- Quinine & quinidine is mainly obtain from the bark, stem & roots of "Cinchona officinalis", "Cinchona ledgeriana", "Cinchona succirubra", "Cinchona calisaya", family - "Rubiaceae".
- Quinine & quinidine are the stereo-isomers which comes under the category of quinine alkaloids.
- Quinine is levo-rotatory-
- Quinidine is dextro-rotatory-

Uses- Quinine is an anti-malarial drugs-

- Quinidine is an anti-arrhythmic drugs-

* Quinine also consider as a better standard-

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iii) Extraction & Isolation -

- Take the dry cinchona bark & powder it with the help of grinder & powder passed through #20
- ↓
- mix the powder with 60% CaOH (slaked lime) & 40% water
- ↓
- keep it for some times during the mixing slaked lime fix the organic acid & Ca^{2+} salt (keep it 24 hrs)
- ↓
- Take the mass & packed in a soxhlet & extract with benzene at room temp.
- ↓
- Collect the benzene extract & filter it, - after conc. & filter (To the conc. extract & sodium carbonate soln to neutralization act \rightarrow pH 6.5)

↓
Precipitate quinine sulphate

↓
Filtrate

↓
• If coloured soln

↓
• If colourless soln

↓
• To the filtrate add Na_2CO_3 & $NaOH$ (1:1)

↓
• Then carbon black is add.

↓
• Which is purified boiling with hot H_2O

↓
• Precipitation take place know extract this soln with diethyl ether

↓
• filter it & conc. the filter

↓
• Quinine sulphate & dissolve & conc.

↓
• To separate quinine & cinchonidine

↓
• keep it for some times quinine sulphate appearance

↓
• keep it for some times, crystal of quinine sulphate app.

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iv) Characterisation / Identification / Chemical test / qualitative

- a). Fall Thalleoquin test-
- b). Ferrocyanide test-
- c). Acid test-
- d). Erythroquin test-

a). Thalleoquin test-

- To the sample add bromine water & ammonia
⇓
- Bright green colour fluorescence appearance.

b). Ferrocyanide test-

- To the sample add bromine water & chloroform
⇓
- Shake it & allow for few minutes
⇓
- Add 1% potassium ferrocyanide solution & 3ml of 5M/5M NaOH solution

Quinine

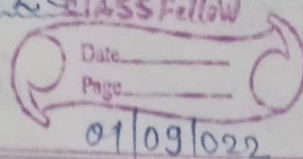
- ⇓
- Chloroform layer turns colourless

Quinidine

- ⇓
- Chloroform layer turns red colour

* R_f -value only for determine in TLC

* HPLC & HPTLC in determine R_f



c) Acid test:-

- To the sample add dilute H_2SO_4 solution
↓
- Sample give blue fluorescence indicate the presence of quinine.-

d) Erythroquin test-

- To the sample add dilute H_2SO_4 solution -
↓
- To the acidic sample add bromine water & shake it well
↓
- Add few drops of strong ammonia solution & neutralized with dilute H_2SO_4 -
↓
- It will gives blue colouration
↓
- Now continuous add dilute H_2SO_4 , it will changes the colour from blue to red.-

v) Analysis - / Quantitative test / characteristics -

a) TLC (Thin layer chromatography) -

b) Spectrometric estimation - / HPLC -

c) HPTLC (High performance thin layer chromatography)

a) TLC - i) sample preparation

ii) stationary phase

iii) solvent system (mobile phase)

iv) spring reagent (detecting reagent) -

v) R_f -value - (Retardation factor) -

* Blank - These are substances/agent in drugs analysis
(other than drugs)

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Dissolve

i) Sample preparation - \uparrow 1mg of quinine in 1ml of methanol

ii) Stationary phase \rightarrow Silica-gel-G (*G = Gypsum)

iii) Solvent phase/mobile phase - Chloroform : Diethylamine
(9 : 1 or 9.6 : 0.4)

iv) Spring reagent/detecting agents -

- Dragendorff's reagent -
- modified dragendorff's reagent (Kraut's reagent) -

v) R_f - value - Quinine = 0.74 - , Cinchonine = 0.66 -
- Quinidine = 0.62 - , Cinchonidine = 0.54 -

⑥ spectrometric estimation - / HPLC (High performance) -

i) Column -

ii) solvent/mobile phase -

iii) Wave length -

iv) Flow rate -

v) R_t (Retention time) -

i) Column \rightarrow microsorb C₁₈

ii) solvent/mobile phase \rightarrow chloroform : Diethylamine -

iii) Wave length - 380nm -

iv) Flow rate - 1ml/minutes -

v) R_t \rightarrow -

① HPTLC - [High performance thin layer chromatography] -

i> Plate -

ii> mobile/solvent phase -

iii> wave length \Rightarrow 380nm -

iv> R_f -

i> Plate - Precoated silica gel 60F₂₅₊ \rightarrow [f = Fluorescence]

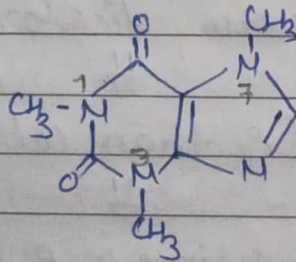
ii> mobile phase - Ethyl acetate : Diethylamine

iii> Flow rate - 1ml/minutes -

iv> R_f - 0.19 minutes -

Caffeine

1> Structure -



2> Chemico-taxonomical distribution -

• Caffeine is a purine alkaloids obtained from 'Tea leaves, coffee seeds, cocoa & other species' -

* It consists of dried leaves of plant known as "Thea sinensis", family - "Theaceae" -

* It is chemically 1,3,7-trimethyl xanthine. It is isolated from tea & coffee seeds during "decaffeination process" -

- * Tea leaves contains 1-4% of Caffeine & Coffee contains 1-2% of Caffeine.
- * It is white powder or white, glistening needle, odourless, bitter in taste, soluble in hot water.
- * Caffeine is a CNS stimulant & diuretic.

3) # Extraction & Isolation -

- Take 20gm of tea leaves & add 10gm of Na_2CO_3 & water
⇓
- Boil for 20-25 minutes on heating mantle & then filter it.
⇓
- Filtrate transfer into separating funnel & add 10 ml of chloroform / dichloromethane
⇓
- Agitation/shake it & then collect bottom transparent layer in beaker -
⇓
- Repeat it for 3-4 times & collect all transparent layer
⇓
- Transfer it into petridish & cover it with aluminium foil.
⇓
- After complete evaporation needle shape crystals of ~~caffe~~ Caffeine obtaine. (white powder)
⇓
- It is recrystallized with alcohol -

4) # Characterisation / Identification / Chemical test / Qualitative

- i) Dragendorff's reagent test
- ii) Murexide test -

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* Murexide test -

- To the caffeine add HCl & potassium chlorate. ($KClO_3$) \Downarrow
- Heated to dryness - \Downarrow
- A purple colour is obtained by exposing the residue to vapours of dilute ammonia -

5> # Analysis / quantitative test -

a> TLC -

- i> Sample preparation - Dissolve 1mg of caffeine in 1ml of methanol / chloroform -
- ii> Stationary phase - silica gel - G
- iii> Solvent / mobile phase - Ethyl acetate : methanol : 4AA [80 : 10 : 10]
- iv> Spiking reagent - Expose to vapour of iodine -
- v> R_f - 0.41 -

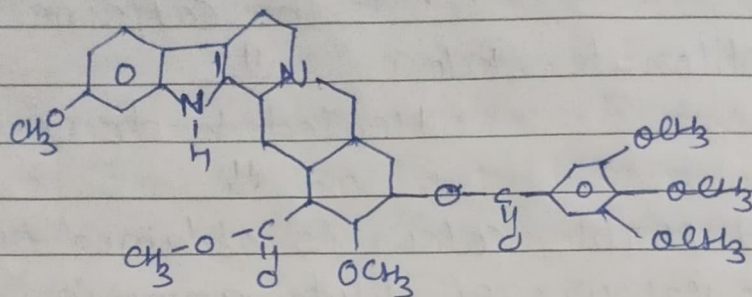
b> HPLC -

- i> Column - microsorb C18 -
- ii> mobile phase - water : methanol (70 : 30) -
- iii> Flow rate - 1ml / minutes -
- iv> Wave length - 275nm -
- v> R_t - 2.6 minutes -

c> HPTLC -

- i> Plate - Pre coated silica gel 60F254 -
- ii> mobile phase - Ethyl acetate : methanol (80 : 20)

- iii> Wave length - 275nm
- iv> R_t - 0.39 minutes -

1> # Structure-2> # Chemotaxonomical distribution-

- Reserpine is an indole alkaloid obtained from the roots of "Rauwolfia serpentina", family-Apocynaceae.
- It is a white or pale buff to slightly yellow, odourless, crystalline powder.-
- It is soluble in alcohol, acetone & chloroform.-
- Reserpine is an anti-hypertensive & anti-psychotic agents.

3> # Extraction & Isolation -

- Rauwolfia root powder is exhaustively extracted with 90% alcohol by percolation -
 ↓
- The alcoholic extract is conc. & dried under reduced pressure below 60°C to yield rauwolfia dry extract
 ↓
- Rauwolfia root dry extract is extracted with Ether, chloroform & 90% alcohol (20:8:2.5) -
 ↓
- Collect the extraction & add little dilute

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ammonia with intermittent shaking. - Add water & allow the drug to settle after vigorous shaking



- Filter off the solution & extract the residue with 4-volume of 0.5N ammonium sulphate in separating funnel. Combine all the extracts.



- The extract is made alkaline with dilute ammonia to liberate alkaloid. Finally it is extracted with Chloroform.



- Collect the chloroform extract, conc. & evaporate on water bath to yield total rauwolfia alkaloids (30-different components) -



- Residue is subjected to column chromatographic fraction for the separation of reserpine.

4> # Identification test -

i> Dragendorff's reagent test -

ii> Reserpine solution + dilute H_2SO_4 & expose to light



- It will give yellow colour with fluorescence

iii> Sample solution + Vanillin in acidic acid



- Violet red colour is produce -

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5) # Analysis -

A) TLC - i) solution & sample preparation - Dissolve 1mg of reserpine in 1ml of methanol & chloroform. -

ii) Stationary phase - silica gel - G -

iii) Solvent phase / mobile phase - Chloroform : Acetone : Diethylamine (50 : 40 : 10) -

iv) Spiking reagent - Dragendorff's reagent -

v) R_f - 0.72

B) HPLC -

i) Column - Reversed phase column -

ii) Flow rate - 1ml/minutes -

iii) wave length - 254nm -

iv) R_t - 21.68 minutes -

v) mobile phase - chloroform : Toluene : Ethyl acetate : Diethylamine (40 : 20 : 20 : 20)

C) HPTLC -

i) Plate - Precoated silica gel G-60F₂₅₄ -

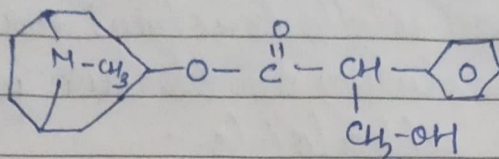
ii) mobile phase - chloroform : Toluene : Ethyl acetate : Diethylamine (40 : 20 : 20 : 20)

iii) wave length - 254nm

iv) R_t - 0.36 minutes -

ATROPINE

1. # structure-



2. # Chemotaxonomical distribution-

• Atropine is a

tropane alkaloid.-

- It is obtained from "atropa belladonna", datuna & stramonium & hyoscyamus niger, family-solanaceae.
- It is a white crystal or crystalline powder & less soluble in water & highly soluble in organic solvents.-
- It mainly act as CNS stimulents & depression action on nervous ending of secretory glands.
- Injection of atropine is used in treatment of bradycardia.-

#3. # Extraction & Isolation-

- Take weighed quantity of coarse powder & moisten with Mascos solution ↓
- Extract the blended mixture in petroleum ether. filter the petroleum ether extract- ↓
- Extract the filtrate with aqueous acetic acid (alkaloids extracted in aqueous layer) - ↓
- Extract the aqueous fraction with solvent ether & separate both fraction. * Discard solvent ether fraction. ↓



* Aqueous (acidic fraction) made alkaline with Na_2CO_3 solution to obtain precipitates (ppt) of tropane alkaloids.-



• Filter the precipitate & dry to obtain residue.-



• Dissolve the residue in diethyl ether



• Filter & conc. the filtrate, atropine crystals will be separated out.-

4) # Identification test-

i. Dragendorff's reagent test-
ii. Clitalin - marin test-

• Take small quantity of the solid atropine & add 2-3 drops of conc. nitric acid (HNO_3) in an evaporating dish & evaporated to dryness on water bath.-



• Then dissolve the residue in 1ml of acetone.
• Add few drops of freshly prepared alcoholic potassium hydroxide solution-



• Violet colouration takes place due to tropane nucleus-

5.2# Analysis-

a) TLC-

i) Sample preparation- Dissolve 1mg of atropine in 1ml of

organic solvents-

ii) Stationary phase- silica gel-4-

iii) Solvent phase- Toluene : Ethylacetate : Diethylamine
(70 : 20 : 10)

iv) Spising reagent- Dragendorff's reagent

* Produce yellow-orange colour spot-

v) R_f - value- 0.7-

b) HPLC- i) Column- C₁₈

ii) solvent/mobile phase- Ortho-phosphoric acid : Aceto-
(70:30) nitrate

iii) Flow rate- 1.4 ml/minutes-

iv) Wave length- 215nm-

v) R_t - 6.6 minutes-

c) HPTLC-

i) Plate- Precoated silica gel-4- 60F254-

ii) mobile phase- chloroform : methanol (70:30)

iii) Wave length- 200nm-

iv) R_t - 0.42 minutes-

Resin

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Date

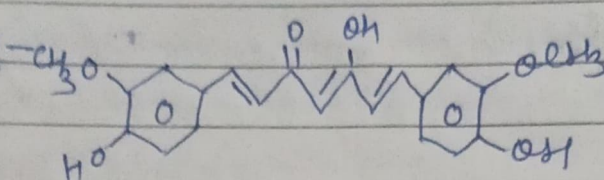
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Curcumin

1>

Structure



[$C_{21}H_{20}O_6$]
[Enol form]

2>

Chemotaxonomical distribution -

• Curcumin or curcuminoids are the diaryl heptoid compd. obtained from the dried rhizomes of turmeric "Curcuma longa", family - 'Zingiberaceae'

- Curcumin is the major colouring principle -
- It is a mixture of curcumin, mono-desmethoxy-curcumin & bis-des-methoxycurcumin. -
- It is an orange yellow, crystalline powder
- Insoluble in water & ether but soluble in alcohol.
- It is used as wound healing, anti-inflammatory, anti-arthritis & anti microbial activities -
- used against peptic ulcer -

3>

Extraction & Isolation

- Curcumin can be obtained by different process -
i.e. Turmeric powder is extracted with alcohol in soxhlet extractor. \Downarrow
- Alcoholic extract is concentrated under reduced pressure & dried -

ii) Powder root is extracted with alcohol in Soxhlet for 6 hrs. \Downarrow

• Collect the extract & again extract hexene \Downarrow

• Collect the hexene extract & re-extract with methanol \Downarrow

• Filter the solution & conc. on water bath to dryness \Downarrow

• To the residue add toluene & NaOH \Downarrow

• Collect aqueous layer from the solution & add acid for gain pH 3.0 & made extract yellow \Downarrow

• Collect the extract & again extracted with diethyl ether \Downarrow

• Collect the ether layer wash with water & dried over MgSO_4 \Downarrow

• Yellow colour curcumin is often.

#4) Identification test-

i) Powder drugs + $\text{H}_2\text{SO}_4 \Rightarrow$ Crimson red colour

ii) Powder drugs + alkaline solution \Downarrow

Red-violet colour -

5. > Analysts -(A) TLC -

- i> Sample preparation \rightarrow Dissolve 1mg curcumin in 1ml of methanol
- ii> Stationary phase \rightarrow Silica-gel-G-
- iii> Solvent phase \rightarrow Chloroform : Ethanol : Glacial acetic acid
(94 : 5 : 1)
- iv> Spising reagent \rightarrow Observed under U.V. light at 366 nm
- v> R_f value - 0.79 -

(B) HPLC -

- i> Column - C18
- ii> mobile phase - Acetonitrile : Acetic acid : water
(80 : 10 : 10)
- iii> Flow rate - 1ml/minutes -
- iv> Wave length - 425 nm -
- v> R_t - 7.04 minutes -

(C) HPTLC -

- i> Plate - Precoated silica gel GF₂₅₄ -
- ii> mobile phase - Dichloromethane : methanol
- iii> wave (50 : 50)
- Wave length - 425 nm -
- iv> R_t - 0.43 -

- "Indian podophyllum" -
OR

Podophyllotoxin

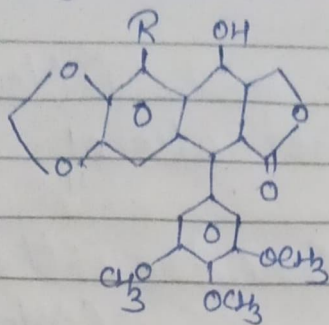
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1. Structure-



R = H (Podophyllotoxin)

R = -OCH₃ (5-methoxy-podophyllotoxin)

2. Chemotaxonomical distribution-

• It is also known as :

Indian ~~phy~~ podophyllum -

* It consists of the dried rhizome & roots of "Podophyllum emodi" (Indian), "Podophyllum peltatum" (American), family - "Berberidaceae"

* Podophyllum potent anti-cancer agents, anti-tumor agents, potent topical anti-viral & used in the HPV (Human papilloma-virus) infection & remove warts.

3. Extraction & Isolation-

• 120gm root powder extract with 300ml methanol in Soxhlet for 6 hrs



• Filter the extract & filtrate concentrate & add 200ml water containing 2ml HCl & cool it.



• Allow mixture to stand for 2 hrs below 5°C & filter it



• Filtrate wash again with acidified water & filter it



- The residue after filtration is mix with sufficient quantity of hot alcohol -



- Filter it & Conc. the filtrate to get podophyllum - (Podophyllotoxin)

4> # Identification test -

- 0.5gm of the drug with 10ml of alcohol & filter
- ↓
- Add 0.5ml copper acetate \Rightarrow Brown ppt's produces
 - Alcoholic extract + Add 5ml of 1M KOH \Rightarrow Stiff jelly is produce-

5> # Analysis -

(A) HPLC - i> Column = Reverse phase C₁₈ -

ii> mobile phase = methanol : water (60:40) -

iii> wave length = 280nm -

iv> Flow rate = 0.9 ml/min. -

v> R_t = 9.45 minutes -

(B) HPTLC -

i> Plate \rightarrow Precoated silica gel 60F₂₅₄ -

ii> mobile phase = Acetonitrile : water (70:30)

iii> wave length = 283 nm -

iv> R_t = 0.41 minutes -

Glycosides

CLASS FOLLOW

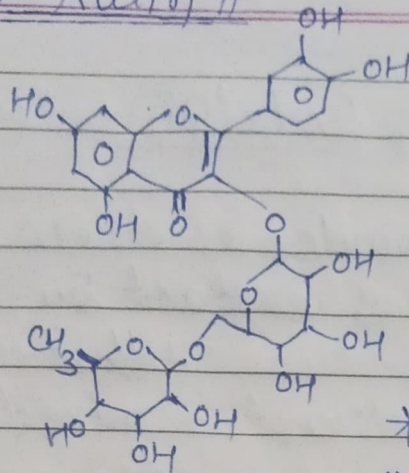
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Rutin

1. # Structure -



Rutin (Rutoside) *

It is called vitamin P -

**
* Pure rutin is yellow or yellow green color, needle shape crystal.

2. # Chemotaxonomical distribution -

It is a citrus flavonoid glycoside containing drugs which mainly obtained from the root, rhizome, leaves part of the plants -

- * The genus of rutin it is very large consists of approximate to 550-750 species & is divided into 11 sub-genera -
- * The rutin is mainly obtained from "Phyllanthus amarus", "Phyllanthus niruri", "Phyllanthus emblica", "Phyllanthus winaria" -
family - Rutaceae

* Rutin is a pale yellow, needle shape structure, present in powder form -

- It is tasteless & odourless -
- Mainly soluble in formaldehyde, pyridine, methanol & alkaline solution -
- Rutin is mainly used as diuretic, bitter astringents, liver, kidney & spleen problem & also act as anti-septic -

** Rutin is also known as "Bhui-aml" -

3> # Extraction & Isolation-

- Take 20gm powder of drugs & add 250ml of 80% ethanol & extract by Soxhletion method
⇓
- Filter the extract & mixed with 25ml of water
⇓
- Again extracted with petroleum ether & chloroform.
⇓
- Take aq. layer & keep it cold for 72 hrs & yellow ppt was separated.-
⇓
- Yellow ppt was wash with chloroform: ethyl acetate: ethanol (50:25:25)
⇓
- After washing ppt is dissolve in hot methanol & filter it
⇓
- The filtrate it come to dryness-
⇓
- Yellow needle shape crystals are obtain-
(Rutin)

4> # Identification test-

- i> Drugs with FeCl₃ ⇒ Give dark green color-
- ii> Drugs with lead acetate ⇒ Orange yellow ppt-
- iii> Drugs with ammonium molybdate & antimony trichloride ⇒ Orange-yellow ppt-
- iv> Schinoda test - few fragments of mg-silbbon + Conc. HCl & add ethanolic extract
⇓
Red to pink colour-

Analysis -(A) HPLC -

- i) Column - Reverse phase C₁₈ -
- ii) mobile phase - methanol : water (70 : 30) -
- iii) Wave length - 280 nm -
- iv) Flow rate - 0.5 ml/min -
- v) R_t - 0.5 minutes -

(B) HPTLC -

- i) plate - precoated silica gel 60F₂₅₄ -
- ii) mobile phase - Acetonitrile : Water (60 : 40) -
- iii) Wave length - 254 nm -
- iv) Flow rate - 5 ml/minutes -
- v) R_f - 0.65 minutes -

(C) IR- value / IR- spectroscopy -

functional group

Peak

-OH

3330

-C=O

1660

-CH

2920

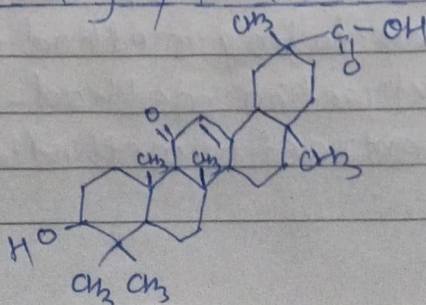
-C=C

1600

-C-O-C

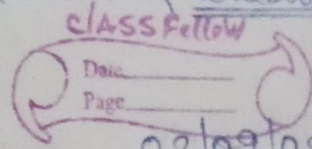
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Exam-

Glycyrrhetinic acid # (5)1) # Structure :-C₃₀ H₄₆ O₄

* Glycyrrhizin is 30-50 times as sweet as sucrose -

* Pure glycyrrhizin is odorless -



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2> # Chemotaxonomical distribution -

• Glycyrrhetic acid is mainly obtained from sweet wood, liquorice & multhi, yastimadhu -

* It is obtained from the dried roots & stolons of "Glycyrrhiza glabra", family - Leguminosae -

* Chief constituent of liquorice is "glycyrrhizin" -

* Glycyrrhizin -

• A triterpenoid saponin glycoside -

• Also called as 'glycyrrhizic A' or glycyrrhizic acid -

* Being a glycoside, glycyrrhizic acid on hydrolysis gives an aglycone & a glycone portions

⇒ Glycone = Glycyrrhetic acid -

⇒ Aglycone = triterpenoidal structure of glucuronic A -

3> # Extraction & Isolation -

• The isolation of glycyrrhizin from glycyrrhiza is based on solubility -

• The three (3) methods of isolation are -

a) Acid precipitating method -

b) Alcohol extraction method -

c) Ammonia extraction method -

a. Acid precipitating method -

- Weigh liquorice powder & add water in the beaker & boil with continuous stirring -



- Decant/filter the supernatant liquid



- filter the remaining residue & collect the filtrate



- Adjust pH 2.8 by the addition of acid -



- Glycyrrhizin precipitates out -



- filter & collect the glycyrrhizin ppt -



- Wash the ppt with cold water to make it free from acid



- Transfer the ppt to china dish & heat gently to remove the water content, shiny brown mass of glycyrrhizin is seen. -

b. Alcohol extraction method -

- Weigh liquorice powder & add 100ml methanol in 500ml beaker & properly mix it



- Add another 100ml methanol & left for 24 hrs -



- filter & collect the filtrate -





- Extract this methanolic extract with 3- portions of petroleum ether, subsequently with benzene, ethylacetate chloroform & solvent ether.

↓ filter

- Transfer methanolic layer into china dish & evaporate on water bath to get glycerzhizin.

c2. Ammonium extraction method-

- Glycerzhiza is extracted with hot water & filtered

↓

- filtrate is acidified with Conc. H_2SO_4 & pH 2.8

↓

- ppt is dissolved in dil. NH_4OH & poured into acetone to ppt ammonium glycerzhizinate-

↓

- The ppt is dissolved in hot water & evaporate to get ammonium glycerzhizinate-

4> # Identification test-

i> foam test-

ii> Haemolysis test-

iii> Liebermann test→

- Drug's solution + Acet 2 anhydride

↓ Boil

- Add H_2SO_4 —————> Blue colour appears

5) # Analysis -a) TLC -

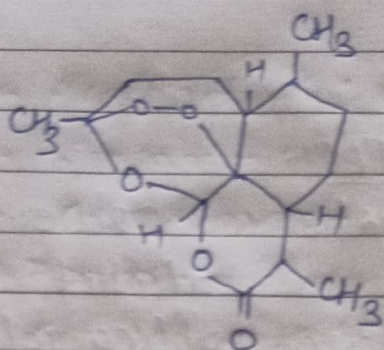
- i) Sample preparation - Dissolve 1mg of liquorice in 1ml of alcohol
- ii) Stationary phase - Silica gel - G-
- iii) mobile phase - ~~form~~ Ethyl acetate : formic acid : GAA (15:1:2)
- Butanol : GAA : water (7:1:2) -
- iv) Spising reagent - Libermann-reagent
- Vapour of Iodine -
- v) R_f - 0.69 -

b) HPCC - i) Column - Reversed phase C18 -

- ii) Spising reagent - photo-diode array detector
- iii) mobile phase - Acetonitrile : phosphoric acid (80:20) -
- iv) Wave length - 230nm -
- v) flow rate - 0.6ml/minutes -
- vi) R_t - 0.6 minutes -

c) HPTLC - i) Plate - Precoated Silica gel 60F₂₅₄ -

- ii) mobile phase - Ethyl acetate : Ethanol : water : Ammonia (60:20:10:10)
- iii) Wave length - 254nm -
- iv) R_t - 0.42 minutes -

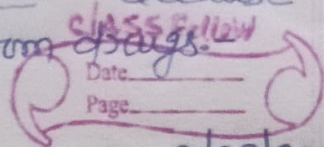
Artemisinin #1. # Structure -2. # Chemotaxonomical distribution -

- Artemisinin is a sesquiterpenoid lactone, obtained from the flower, head of part of the plant "Artemisia annua", "Artemisia citra", family - Compositae/Asteraceae.
- * Synonyms of artemisinin is "Santonin" -
- * Artemisinin is a white crystalline powder, soluble in organic solvents & insoluble in water -
- * Artemisinin is an anti-malarial drug which used in the treatment of malarial & many other disease.

3. # Extraction & Isolation -

- The leaves are air dried, coarsely powdered & extracted with petroleum ether
⇓
- The extract is conc., dried & re-dissolved in chloroform. Add acetonitrile to ppt sugar & waxes -
⇓
- Filter & collect the filtrate. Evaporate to dryness to yield residue. -
⇓
- The chromatographic fractionation of the conc. on silica gel by eluting with chloroform: ethyl acetate

* Primary solvent \Rightarrow "Petroleum ether" \rightarrow Because remove the impurity from drugs.



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yields the fraction of artemisin. -



- The fraction containing artemesin could be crystalline from cyclohexane or 50% ethanol. -

4. # Identification test -

- Boil 1gm finely powdered drug with 10ml alcohol & filter.



- To the filtrate, add NaOH & heat again. -



- The liquid develops red colour -



- Indicate the presence of artemisin. -

5. # Analysts -

(A) T.L.C. - i. Sample preparation - Dissolved 1mg of artemisin in 1ml of chloroform -

ii. stationary phase - silica gel - G -

iii. mobile phase - Petroleum ether : Ethyl acetate (5:5) -

iv. spraying reagent - Para-dimethyl-amino-benzaldehyde & at 80°C to produce colour -

v. R_f value - Compare with standard artemisin -

(B) HPLC -

i. Column - Reversed phase C₁₈ -

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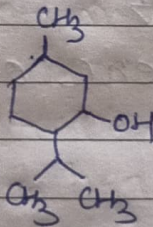
- ii) mobile phase - Acetonitrile : water (80:20) -
- iii) wave length - 250 nm -
- iv) Flow rate - 0.5 ml/min -
- v) R_t - 0.7 min -

(C) HPTLC -

- i) Plate - Precoated silica gel 60F254 -
- ii) mobile phase - n -hexane : Ethyl acetate (70:30) -
- iii) wave length - 540 nm -
- iv) R_t - 0.7 min -

Menthol

1) # Structure -



2) # Chemotaxonomical distribution -

- menthol is an organic compd., belong to terpenoids group -
- * menthol is mainly obtained from leaves part of the plant "Mentha piperita", "Mentha spicata", family - "Labiatae" -
- * menthol is a waxy, crystalline, clear substance or white in colour -
- * menthol is solid at room temperature -
- * menthol is a mono-terpene alcohol obtained from different variety of mint oils or peppermint oils -

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- * Freely soluble in organic solvents like alcohol, chloroform, Carbon tetrachloride, ether & other solvent -
- * menthol is mainly used as local anesthetic & counter irritant. -
- * menthol also used in throat irritation & dental care as a topical anti-bacterial agents -

Extraction & Isolation -

- Take the accurately weighed quantity of coarse powder of mentha piperita parts just before flowering
↓
- Extract the peppermint oil by water distillation method by the help of Clevenger apparatus -
↓
- Separate the oil & allow cooling. Crystals of (-) menthol will separate out. -
↓
- Collect the crystals by "Centrifugation" -
↓
- Re-crystallize menthol from acetone or any other low boiling point solvent. -

Identification test -

- few gm of menthol + few drops of conc. H_2SO_4 + few drops ~~water~~ Vanillin Sulfuric acid reagent
↓
- Orange colour appear \Rightarrow when add water in orange colour ppt \Rightarrow orange change violet

* Small piece of KOH added into test tube containing 1ml oil, then heated \Downarrow

- Cool the solution & add 1ml of diethyl ether & few drops of carbon disulfide

\Downarrow

- Yellow residue is obtain -

5) # Analysis -

(A) T.L.C. - \Rightarrow Sample preparation -

in 1ml of menthanol -

• Dissolved 1mg of menthol

ii) Stationary phase - Silica gel - G -

iii) mobile phase - Pure chloroform -

iv) Spising reagent - 1% Vanillin - Sulphuric acid reagent
& heat the plate 110°C for 10 minutes

v) R_f-value - 0.48 - 0.62 -

(B) HPLC -

i) Column - Shim pack VP - ODS -

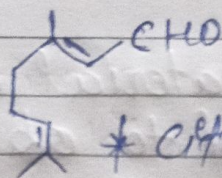
ii) Column temp. - 25°C

iii) mobile phase - Pure ethyl acetate -

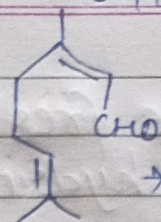
iv) Wave length - 322 nm -

v) R_t \rightarrow 0.4 min. -

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Citral ## Structure -

* Citral-A



* Citral-B

Chemotaxonomical distribution -

Citral is also known as "Lemonal"

* Citral is monoterpene aldehyde found in variety of sources like- lemon grass, lemon peels & orange peels etc.

* Citral is mainly obtain from the lemon grass "Cymbopogon flexuosus", family- Graminae -

* 75-85% of citral present in lemon grass -

** Citral have -2- isomers -

i. E-isomers is known as "Geranial / Citral-A" -

ii. Z-isomers is known as "Neral / Citral-B" -

* It is yellow colour off mainly soluble in organic solvent & insoluble in water. -

** Geranial & Neral both are light oily liquids with lemon odour -

* Citral is mainly used in perfume, flavouring agents & many other chemical industry. -

** Citral also have significant production of Vit. A -

* Citral has strong anti-microbial properties -

* It has an effect on insect & act as mflr repellent for some species. -

Extraction & isolation -

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- The fresh plant material is collected & subjected to hydro-distilled to obtain lemon grass oil
- ⇓
- Purification of lemon grass oil is performed by fractional crystallization -
- ⇓
- To the total oil, first sodium sulphite is added. The citral gets converted into its sulphite salt -
- ⇓
- The salt crystallizes out of the solution -
- ⇓
- The crystals are filtered & washed with ether or chloroform -
- ⇓
- The product is then subjected to sodium carbonate treatment to recover citral -

4. # Identification test -

i) Sudan red III -

- Take a thin section of drug & add alcoholic solution of sudan red III
- ⇓
- Red colour, which indicates presence of volatile oil -

ii) Tincture alkane -

- Take thin section of the drug & add a drop of tincture alkane -
- ⇓
- Red colour is produced which indicates presence of volatile oil -

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> # Analysis -(A) TLC.

- i) Sample preparation - Dissolve 1mg of citral in 1ml of ~~menthol~~ - methanol.
- ii) Stationary phase - Silica gel - 4.
- iii) mobile phase - Pure chloroform -
- iv) Spising reagent - 2,4-dinitrophenyl hydrazine reagent
- v) Colour - Yellow to orange -
- vi) R_f value - 0.51 -

(B) HPLC - i) Column - ODS-hypersill column -

- ii) mobile phase - methanol -
- iii) Flow rate - 1ml/minutes -
- iv) Wave length - 254 nm -

(C) HPTLC -

- i) plate - Pre coated silica gel 60f254 -
- ii) mobile phase - Pure toluene -
- iii) wave length - 595 nm -
- iv) Flow rate - 1ml/min. -
- v) R_f - 0.7 min. -

** Screening speed = 20 mm/sec. -